

1 Population dynamics of Amazonian floodplain forest species support spatial variation on genetic
2 diversity but not range expansions through time.

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14

15 **Abstract**

16 **Aim:** We tested if historical demographic changes of populations occurring on the floodplains of
17 a major Amazon Basin tributary could be associated with range expansions from upper and
18 middle sections of the river, following the establishment of widespread river-created
19 environments during the Late Pleistocene and Holocene.

20 **Location:** Solimoes River, Western Amazon, South America

21 **Taxon:** *Myrmoborus lugubris*, *Thamnophilus cryptoleucus* and *Myrmotherula assimilis*

22 **Methods:** We analyzed thousands of UltraConserved Elements to explore spatial patterns of
23 genetic diversity and connectivity between individuals. Range expansions were tested with
24 alternative methods. We quantified habitat preference for the analyzed species in order to test if
25 the occupation of dynamic habitats could predict spatial patterns of genetic diversity.

26 **Results:** Our study did not support shared population range expansions related to historical
27 regionalized changes in habitat availability. We found considerable variation in the spatial
28 distribution of the genetic diversity between studied taxa, and that species with higher levels of
29 specialization to dynamic environments have a more heterogeneous distribution of genetic
30 diversity and reduced levels of gene flow across space.

31 **Main conclusions:** Our results suggest that demographic expansions along the Solimões River
32 might be linked to geographic homogeneous oscillation in the distribution of floodplain
33 environments, promoting effective population size changes but not range expansion. We found
34 that habitat specificity might be a good predictor of population connectivity along the
35 Amazonian floodplains.

36

37 **Introduction**

38 Understanding the factors that shape the spatial distribution of species is a long-standing
39 challenge in comparative biology that is attributable to the complex interaction of environmental
40 and species trait variables. Over evolutionary time scales the geographic distribution of
41 populations might be affected by physiographic processes (Silva et al., 2019; Peter et al., 2020),
42 climatic oscillations (Hewitt, 2000; Raposo do Amaral et al., 2018; Musher et al., 2020), biotic
43 interactions (Araújo & Luoto, 2007; Wisz et al., 2013), and trait evolution (Alves et al., 2019),
44 driving dispersion, extinction, and cladogenesis. Although processes such as the appearance of
45 novel traits, interspecific competition, and colonization of new environments might lead to a
46 single burst of range size change, in many cases species distributions are dynamic over time,
47 oscillating between periods of range contraction followed by expansion (Davis & Shaw, 2001;
48 Taberlet & Cheddadi, 2002; Carnaval & Moritz, 2008). Range expansions are expected to be
49 accompanied by a rapid increase in the number of individuals, and strong spatial directionality
50 (Excoffier et al., 2009). At the expansion front, successive founder events lead to a fast
51 differentiation from the source, producing a characteristic demographic signature that is
52 distinguishable from non-spatial population growth (Excoffier et al., 2009; François et al., 2010;
53 Peter & Slatkin, 2013; Alvarado-Serrano & Hickerson, 2018). By tracking the spatial dynamics
54 of populations one can locate zones of climatic stability (Potter et al., 2016), track sources of
55 population invasions (Fischer et al., 2017), and understand how genetic diversity was affected by
56 past environmental changes (Reid et al., 2019), allowing for more robust predictions to explore
57 classical diversification hypotheses.

58 One of the main processes used by phylogeographic approaches to explain population
59 size changes through time is the variation on available habitat distribution due to vegetation
60 response to historical climate change, as described by the refugia hypothesis (Haffer, 1969, 1997;

61 Hewitt, 2004). This mechanism suggests that populations tend to be isolated in "islands" of
62 optimum habitats during glacial (e.g., lowland forest taxa) or interglacial (e.g., montane forest
63 taxa) periods followed by population expansion and secondary contact as habitats expand. In the
64 Neotropics, the region for which the refugia hypothesis was initially proposed (Haffer, 1969), the
65 effects of past climatic oscillations have been used to explain patterns of genetic diversity in
66 multiple environments, including the Atlantic Forest (Carnaval et al., 2009; Amaral et al., 2018;
67 Silva et al., 2019), Amazonia (Thom et al., 2020b), Caatinga (Gehara et al., 2017), the Chaco dry
68 forests (Camps et al., 2018) and the Andes/Patagonia (Cosacov et al., 2010). However, few
69 studies have sought to understand if demographic patterns are linked to range size change
70 (Camargo et al., 2013; Guarnizo et al., 2016; Baranzelli et al., 2017). By testing models
71 including range expansions, one can explicitly explore alternative hypotheses regarding the
72 location of putative stable habitats through time, providing a better understanding of the
73 historical dynamic of specific environments (Potter et al., 2016; He et al., 2017; Reid et al.,
74 2019).

75 In the Amazon floodplains, historical variation in precipitation and global sea-level
76 affected sedimentation dynamics of the main rivers (Latrubesse & Franzinelli, 2005; Irion et al.,
77 2009; Soares et al., 2010; Pupim et al., 2019), potentially shaping the distribution and
78 connectivity of populations occurring on river-created environments (Aleixo, 2006; Choueri et
79 al., 2017; Thom et al., 2020b). Phylogeographic studies have reported demographic expansions
80 for populations of birds specialized in river islands, suggesting widespread habitat alterations
81 across the entire Amazon river system (Aleixo, 2006; Thom et al., 2020b). Historical oscillations
82 in the availability of floodplain environments tend to be more pronounced in regions where large
83 tributaries with distinct sediment loads meet (Gualtieri et al., 2018). For instance, the confluence
84 of the Negro (low sediment load) and Solimoes rivers (high sediment load) leads to an increase
85 in the water discharge and dilution of sediment concentration of the main Amazon river channel,
86 promoting erosion and sediment bypass (Thom et al., 2020b). The current dynamics between
87 Negro and Solimoes rivers prevent the building of long-lasting substrates for flooded habitats
88 downstream of the confluence for approximately 150 km, until the high sediment input of the
89 Madeira River, which restores suitable conditions for the development of large and stable
90 floodplains in the main channel of the Amazon river (Filizola & Guyot, 2009). Recent

91 phylogeographic studies suggest that the central portion of the Amazon Basin, between the
92 Negro and Madeira rivers, is indeed a suture zone, partially or completely isolating recently
93 diverged populations, mostly restricted to Solimoes, Negro, Madeira, and lower Amazonas rivers
94 (Choueri et al., 2017; Thom et al., 2018, 2020b). This spatial congruence indicates that fluvial
95 sedimentation dynamics may directly influence the connectivity of species occurring in river-
96 created environments along Amazonian floodplains through time.

97 The historical interaction between the sediment-poor Negro River and the sediment-rich
98 Solimões-Amazonas main channel, mediated by past oscillations in precipitation and sea level,
99 may have been dynamic over time, shaping the availability of flooded habitats in central
100 Amazonia. For example, the Negro River is currently impacted by a damming effect caused by
101 the larger water input from the Solimões-Amazonas River, maintaining a ria lake at its lower
102 section, which prevents island formation and creates a barrier for the biota associated with river-
103 created habitats (Choueri et al., 2017). Similarly, past increases in the Negro River discharge
104 could have periodically affected the flooding pulse of the Solimões (Irion et al., 2009; Passos et
105 al., 2020), causing suppression of seasonally flooded habitats from its lower and middle sections
106 (Latrubesse & Franzinelli, 2005; Soares et al., 2010; Passos et al., 2020). Under this scenario, we
107 expect seasonally flooded habitats in the upper Solimões River to be historically more stable than
108 areas closer to the Negro River confluence (lower Solimoes River). Hence, the demographic
109 expansions detected by previous studies for populations of birds (Thom et al., 2020b) would be
110 linked to recent range expansions from the upper/middle towards the lower sections of the
111 Solimoes River. This demographic process should lead to higher genetic diversity on upriver
112 populations, and if this was a pervasive mechanism, we expect to detect the same pattern in co-
113 occurring populations that occupy similar habitats. Alternatively, population demographic
114 expansions could be related to geographic homogeneous oscillation in habitat availability (e.g.,
115 oscillations in the carrying capacity of the environments). Under this scenario, we expect the
116 genetic diversity to be mostly explained by the geographic distance between individuals
117 (Isolation by distance), with a relatively homogeneous distribution of the genetic diversity across
118 the whole extension of the Solimoes River, or governed by species-specific characteristics. In the
119 latter, species would have variable patterns across space, potentially associated with habitat
120 specificity.

121 The Amazon floodplains are an intricate mosaic of microhabitats that roughly segregate
122 with the sedimentary dynamic of the rivers and the geographic distance from the main channel
123 (Junk et al., 2012, 2015). Areas next to the margins or on islands of rivers with high sedimentary
124 budgets as the Solimoes, tend to be more dynamic over time, keeping vegetation in early
125 successional stages of regeneration due to the constant erosion and sedimentation. Given the high
126 diversity of microhabitats along the floodplains, we might expect that species with higher
127 dependence on more dynamic environments should have been more affected by historical
128 changes on the floodplains, having a more heterogeneous spatial distribution of genetic diversity.
129 In this case, higher association to more dynamic habitats should predict spatial levels of genetic
130 diversity.

131 We explored the levels of connectivity and the spatial distribution of the genetic diversity
132 of three antbird species, *Myrmoborus lugubris*, *Thamnophilus cryptoleucus*, and *Myrmotherula*
133 *assimilis* with populations restricted to the Solimões River, testing if the population demographic
134 expansions reported in a previous study (Thom et al., 2020b) are related to range expansions
135 following a period of isolation in the upper/middle course of this river. We reanalyzed the data
136 published by Thom and collaborators (2020b) using a set of methods designed to track
137 population range expansions and explore how genetic diversity is distributed across space. Our
138 results refute the idea of congruent range shifts related to historical regionalized changes in
139 habitat availability. Instead, our data suggest that demographic expansions along the Solimões
140 floodplains might be associated with variation in habitat availability across the entire range of the
141 species, promoting effective population size changes but not range expansion. We found that
142 species with higher levels of specialization to dynamic environments have a more heterogeneous
143 distribution of genetic diversity and reduced levels of gene flow across space, indicating that
144 habitat specificity might be a good predictor of population connectivity along the Amazonian
145 floodplains.

146

147 **Material and Methods**

148 We selected samples from the Solimões River populations of *Myrmoborus lugubris* (N =
149 31), *Thamnophilus cryptoleucus* (N = 20), and *Myrmotherula assimilis* (N = 16) that were
150 previously analyzed by Thom et. al (2020b; Figure 1; Table S1). Genome-wide variation was

151 obtained through the sequence capture of Ultraconserved Elements (UCEs) using a probe set
152 targeting 2,312 UCE loci and 97 additional probes targeting exons commonly used in Avian
153 phylogenomic studies (Hackett et al., 2008; Kimball et al., 2009; Faircloth et al., 2012). Raw
154 sequences were downloaded from the Sequence Read Archive (SRA; BioProject ID:
155 PRJNA595086; SubmissionID: SUB6206537; www.ncbi.nlm.nih.gov/bioproject/595086). We
156 used Trinity 2.4 (Grabherr et al., 2011) to assemble *de novo* reads into contigs. The contigs were
157 matched to the probes with LASTZ (<http://www.bx.psu.edu/~rsharris/lastz/>) using
158 “match_contigs_to_probes.py” from PHYLUCE (Faircloth et al., 2012; Faircloth, 2016). Contigs
159 that did not align to probe sequences and those that matched multiple loci were removed. The
160 resulting fasta files from the different individuals were aligned in MAFFT v7.475 (Katoh &
161 Standley, 2013), allowing for missing individuals and without trimming long ragged-ends. These
162 long ragged-ends were then trimmed by applying a threshold of 50% of missing sequences
163 among individuals with TrimAl v1.4.rev15 (Capella-Gutierrez et al., 2009). The longest
164 sequence without indels of each locus was selected as a reference for the following steps. First,
165 reads of each individual were aligned to the reference allowing 4 mismatches per read using
166 BWA v0.7.17-r1188 (Li & Durbin, 2009). The sam files obtained were converted to bam format
167 with Samtools v1.10 (Li et al., 2009). Reads were trimmed to match the reference using
168 CleanSam.jar, reassigned to groups with AddOrReplaceReadGroups.jar, and duplicated reads
169 were identified with Markduplicates.jar from PICARD v.2.0.1 (Broad Institute, Cambridge, MA;
170 <http://broadinstitute.github.io/picard/>). We merged individual bam files into a single bam file of
171 all samples with MergeSamFiles.jar from PICARD. With GATK v3.6 (McKenna et al., 2010) we
172 realigned all reads, identified indels (RealignerTargetCreator; IndelRealigner), and called SNPs
173 hard-masking low-quality bases (< Q30) with UnifiedGenotyper and VariantAnnotator. We
174 obtained raw vcf files for each species that were filtered for a minimum read depth of > 8 using
175 VCFTOOLS v0.1.15 (Danecek et al., 2011). Loci with any site with heterozygosity higher than
176 0.75 were excluded. Finally, we randomly selected one SNP per locus, excluding sites with
177 missing data, resulting in a complete SNP matrix.

178

179 *Genetic Structure*

180 We tested the best-fit number of ancestral populations (k) for each species complex and

181 clustered individuals to populations in sNMF v1.2 (Frichot et al., 2014) by applying a sparse
182 non-negative matrix factorization to compute least-square estimates of ancestry coefficients. We
183 explored values of k between 1 and 8 and performed 100 replicates for each value with an alpha
184 regularization parameter of 100. Given that populations occurring in linear and continuous
185 environments such as rivers are more subjected to the effects of the geographic distance among
186 individuals, and that IBD can generate discrete genetic clusters (Thomaz et al., 2016), we tested
187 genetic structure controlling for the effects of IBD in the R package conStruct v1.0.3 (Bradburd
188 et al., 2018). ConStruct simultaneously estimate continuous and discrete patterns of population
189 structure by assuming a rate of decay in the relatedness among individuals as a function of the
190 geographic distance. Thus, discrete groups are only assumed when genetic variation significantly
191 deviates from an IBD scenario. We ran models assuming the same number of layers as the
192 number of ancestral populations obtained in the best sNMF model and calculated the relative
193 layer contribution with the function “calculate.layer.contribution”, in order to observe if the
194 genetic structure inferred by sNMF could be explained exclusively by IBD. For each conStruct
195 run, we performed 50,000 iterations discarding the first 50% as burn-in. We calculated the
196 geographic distance matrices for each species following river connectivity.

197

198 *Isolation by distance, effective diversity, and gene flow*

199 We identified spatial variation in gene flow among localities and tested deviations from
200 isolation by distance (IBD) scenario for each species with EEMS (Petkova et al., 2016). EEMS
201 estimates the effective migration surface among drawn demes mapping genetic differentiation
202 based on a spatially explicit approach by using a Markov Chain Monte Carlo (MCMC) to
203 estimate effective demographic parameters given the observed genetic dissimilarity between
204 individuals. Euclidian genetic dissimilarity matrices between individuals for each species
205 complex were generated in ADEGENET v2.1.3 (Jombart & Ahmed, 2011). Habitat polygons
206 were produced based on the geographic distribution of each species complex and 300 demes
207 were distributed over the habitat area. Each MCMC run was performed for 30×10^6 generations
208 with the first 5×10^6 generations excluded as burn-in. Maps were generated using additional
209 features of the EEMS R package. We calculated the inbreeding coefficient (F) per individual in
210 VCFTOOLS and interpolated the obtained values across the species distribution in QGIS

211 (<https://www.qgis.org/>). Additionally, to test IBD we performed a Mantel correlation test
212 between genetic and geographic distances in ADEGENET with 999 replicates.

213

214 *Geographic range expansion*

215 We tested for population range expansion using two alternative approaches, first
216 estimating the directionality index using rangeExpansion v0.0.0.9 in R (Peter & Slatkin, 2013,
217 2015) and second, estimating the geographic spectrum of shared alleles in the GSSA v0.0
218 program (Alvarado-Serrano & Hickerson, 2018). The rangeExpansion applies a founder effect
219 algorithm assuming a stepping stone model of population expansion from a single location,
220 testing the strength of the spatial expansion and the most likely origin of the founder effect,
221 measuring the effective founder distance. The effective founder distance represents the size of
222 the deme in which the effective population size is reduced by 1% during the founder event
223 occurring in the expansion front. Thus, low values for the effective founder distance suggest a
224 strong founder effect. For this approach, in order to calculate the derived state of each SNP, we
225 assumed as outgroup one individual from a closely related population (Thom et al., 2020b).

226 The GSSA (Alvarado-Serrano & Hickerson, 2018) captures the range expansion signal
227 by summarizing the shared co-ancestry between sampled localities. This statistic consists of
228 estimating the absolute number of minor alleles shared between sampled localities, taking into
229 account the geographic distance between localities. Given that the amount of genetic drift
230 increases with the distance to the founder event (source of the expansion), the obtained per
231 locality histograms contain information regarding the position of a given locality with respect to
232 the source of the geographic expansion. Some advantages of the GSSA are that no outgroup
233 needs to be used and it can handle less geographic dense sampling when compared to the
234 rangeExpansion method. GSSA implementation allows for the user to provide a geographic
235 distance matrix, instead of assuming linear distances from geographic coordinates. Given the
236 intimate relationship of the studied species with the floodplains, we calculated the geographic
237 distance matrixes for each species following the river's connectivity. The information contained
238 in the shape of the GSSA histogram was summarized using Harpending's raggedness index
239 (Harpending, 1994) and the obtained values per locality were interpolated across the distribution
240 of each species in QGIS. We expect that locations further away from the expansion source share

241 more genetic variants with nearby locations that were colonized through the same expansion
242 route, and thus have higher values for the Harpending's raggedness index. For both analyses, we
243 selected a single individual per locality with the lowest amount of missing data.

244

245 *Habitat occupancy and river dynamism*

246 To quantify differences in habitat occupation between the three species that could predict
247 the spatial patterns of genetic diversity we used a metric of river dynamism extracted from the
248 Occurrence Change Intensity Map available at <https://global-surface-water.appspot.com/> (Pekel
249 et al., 2016). This layer contains information on whether the water surface increased (e.g.
250 erosion) or decreased (e.g. deposition of sediments) around bodies of water, and its intensity
251 based on both the intra and inter-annual variability and changes, extracted from Landsat satellite
252 images between 1984 and 2019. On this layer, values range from -100 to 100, where negative
253 values represent a loss of water surface, positive values represent an increase of water surface,
254 and zero represent stable water surfaces. To calculate the dynamicity of a radius around
255 occurrence records for the studied species, we: 1) converted negative values to positive, so both
256 gain and loss represent how much a given pixel has changed over time; 2) drew a buffer of 2km
257 around every stable body of water, removing areas that were permanently water across the
258 analyzed time section; 3) used random drawn points and occurrence records for the three species
259 to calculate average values of dynamicity using a 1000 meters radius around the coordinates. To
260 create the random distribution we draw 1000 points across the floodplains of the Solimoes Basin.
261 We plotted the values for dynamicity and calculated pairwise Wilcoxon tests in R, to explore if
262 values extracted from random points and each species statistically differ. Spatial layers were
263 processed using the raster v3.4 in R (Hijmans & van Etten, 2013). Occurrence records were
264 obtained from video, photos, and sound recordings from Macaulay Library
265 (www.macaulaylibrary.org) and Xeno-canto (www.xeno-canto.org), and deposited specimens on
266 scientific collections (Table S2). Each record was individually checked, and records with a
267 generic locality description or low-resolution coordinates were excluded. Occurrences at the
268 same locality and date were counted only once. Geographic coordinates outside the 2km buffer
269 were moved to the closest pixel within the buffer.

270

271 Results

272 Our bioinformatics pipeline produced raw vcf files with 5,602 (16.8 mean depth; 2.8 sites
273 per loci), 10,624 (28.4 mean depth; 4.7 sites per loci), and 10,283 (19.9 mean depth; 7.1 sites per
274 loci) variant sites for *M. lugubris*, *T. cryptoleucus*, and *M. assimilis*, respectively. After randomly
275 selecting one SNP per UCE loci we obtained matrices with 1,476, 1,805, and 1,776 SNPs for *M.*
276 *lugubris*, *T. cryptoleucus*, and *M. assimilis*, respectively. The number of genetic clusters obtained
277 with sNMF was partially concordant between species. For *M. lugubris* and *T. cryptoleucus*, the
278 best value of masked cross-entropy was achieved with K=2 while for *M. assimilis* the best value
279 suggested K=1 (Figure 2, S1). However, the results for K=2 for *M. assimilis* recovered a similar
280 geographic structure as the ones observed in the other two species, with a gradual transition over
281 the central portion of the Solimões basin (Figure 2). For *T. cryptoleucus* and *M. assimilis*, this
282 transition in the ancestry coefficients occurred between Tefé and Codajas (localities 9 to 11 in
283 Figure 1) while for *M. lugubris* it occurred more to the west, at Santo Antonio do Ica (locality 6
284 in Figure 1). When accounting for geographic distance with conStruct, assuming 2 layers, the
285 relative contribution of the second layer was less than 1% and without geographic
286 correspondence in all three species. These results suggest that the genetic structure obtained with
287 sNMF is better explained by isolation by distance (IBD).

288 The EEMS detected slightly lower effective migration than the average in the eastern
289 Solimões basin, suggesting progressively more dissimilar individuals towards the east in all three
290 species (Figure 2). However, the estimated effective diversity produced contrasting results
291 between species. While for *M. lugubris* we detected lower diversity in eastern localities, for *T.*
292 *cryptoleucus* there was considerably lower diversity in western localities, and for *M. assimilis* we
293 found a relatively homogeneous distribution of genetic diversity (Figures S2, S3). The results for
294 a Mantel test were congruent with EEMS analyses, significantly supporting an IBD scenario in
295 all three species (p-value < 0.01; Mantel r statistics of 0.59, 0.45, and 0.74 for *M. lugubris*, *T.*
296 *cryptoleucus*, and *M. assimilis*, respectively). The inbreeding coefficient indicated a similar
297 scenario as reported by EEMS effective diversity results for *M. lugubris* and *M. assimilis*, and a
298 homogeneous distribution for *T. cryptoleucus* (Figure S4).

299 The geographic expansion model was strongly supported over an equilibrium/IBD model
300 in the rangeExpansion analysis only for *T. cryptoleucus* (p-value < 0.001; q = 0.0004; Table 1;

301 Figure S5) indicating a geographic expansion from western Solimões. For *M. lugubris* and *M.*
302 *assimilis* the equilibrium/IBD model could not be rejected (p-value of 36.17 and 0.86 for *M.*
303 *lugubris* and *M. assimilis*, respectively). The GSSA estimations support a geographic
304 directionality for the raggedness index in *M. lugubris*, with lower values in the eastern and
305 higher values in the western Solimões as suggested by the rangeExpansion (despite the non-
306 significant p-value; Figure 3). For *T. cryptoleucus*, despite the significant results in the
307 rangeExpansion, there is no geographic signal supporting a range expansion using the GSSA.

308 Data processing of occurrence records produced 60, 92, and 44 points for *M. lugubris*, *T.*
309 *cryptoleucus*, and *M. assimilis*, respectively. The pairwise Wilcoxon tests indicated that all three
310 species occupy areas that are more dynamic than randomly drawn points, with *M. lugubris* and
311 *T. cryptoleucus* occurring in areas significantly more dynamic than *M. assimilis* (Figure 4).

312

313 Discussion

314 *Demographic histories are not linked to a shared pattern of range expansion*

315 We used the genetic variation within thousands of Ultraconserved Elements to explore
316 spatial patterns of genetic diversity in three species of antbirds that inhabit river-created
317 environments along the Solimões River. Previous studies suggested that populations of these
318 three species experienced synchronic demographic expansions in the last 0.2 million years ago
319 (Thom et al., 2020b). Here we tested if demographic expansions were linked to geographic range
320 expansions associated with historical alteration in the distribution of river-created environments
321 (Latrubesse & Franzinelli, 2005; Irion et al., 2009; Soares et al., 2010; Govin et al., 2013; Pupim
322 et al., 2019). Our results supported that only one species, *T. cryptoleucus*, had a significant range
323 expansion with geographic origin in the western Solimoes River. The spatial distribution of the
324 genetic diversity reported here refutes historical events triggering common regionalized changes
325 in population ranges along the Solimoes River. Our results suggest that despite the high
326 dynamism of floodplain environments over time, significant regional suppression of seasonally
327 flooded habitats was probably restricted to specific portions of the floodplains. For example, the
328 central portion of the Amazon, where the sedimentary basin of the Solimões River flows into the
329 eastern Amazon basin, delimits the distribution of many floodplain lineages and is a suture zone
330 splitting populations and species with distinct levels of genetic structure and gene flow (Farias &

331 Hrbek, 2008; Albernaz et al., 2012; Zimmer & Isler, 2016; Thom et al., 2020b). Hence, the
332 demographic expansions reported by Thom and collaborators (2020) might be related to
333 homogeneous oscillations in habitat availability across the entire geographic distribution of the
334 populations, promoting population size changes but not directional range contractions and
335 expansions.

336

337 *Genetic diversity is heterogeneously distributed across the Solimoes floodplains*

338 We observed species-specific patterns for the spatial distribution of genetic diversity
339 along the Solimões River, potentially associated with variation in the occupancy of dynamic
340 habitats (Figures 2, S2, S3, S4). The antbird species studied here are intimately related to the
341 river edge forests that are mostly distributed over islands of large rivers (Remsen & Parker,
342 1983; Junk et al., 2011). Given the low dispersal capacity of the studied species (Zimmer & Isler,
343 2016) and the historical dynamism of river islands, one might expect a shared pattern of genetic
344 diversity between species. However, our data do not support this scenario, with the three species
345 having different patterns of the spatial distribution of their genetic diversity. Despite occupying
346 similar environments, our data suggest that species differ in their levels of association to
347 dynamic habitats. We found that species occurring in significantly more dynamic areas have
348 higher inbreeding coefficients, more heterogeneous spatial distribution of genetic diversity, and
349 more variable levels of effective gene flow across space. These results indicated that habitat
350 specificity might predict the spatial connectivity of floodplain species. Despite the congruency in
351 historical demography, reported in previous studies (Thom et al., 2020b), our results suggest a
352 pronounced effect of intrinsic ecological traits leading to distinct dispersal capacities affecting
353 the spatial distribution of the genetic diversity within populations.

354 Heterogeneous patterns of genetic diversity have been reported along the Amazonian
355 floodplains (Choueri et al., 2017; Jardim de Queiroz et al., 2017; dos Anjos Oliveira et al., 2019;
356 Farias et al., 2019). Choueri and collaborators (2017) studying landscape genetics of river island
357 specialist birds found that genetic diversity over Negro River archipelagos is not homogeneously
358 distributed and that the temporal dynamics of formation and disappearance of islands shaped the
359 genetic structure and gene flow between populations. Additionally, these authors suggest that the
360 increase in sediment accumulation and island formation during the Holocene expanded

361 connectivity between archipelagos (Latrubesse & Franzinelli, 2005; Latrubesse & Stevaux, 2015;
362 Choueri et al., 2017). However, in contrast to the demography of Solimões River populations,
363 only slight signals of expansion were detected for the Negro River populations, corroborating the
364 idea that rivers with black and clear water (sediment-poor) are less dynamic over time than
365 white-water rivers (sediment-rich rivers), such as the Solimões (Thom et al., 2020b).

366 The patterns of genetic structure we detected with sNMF (K=2) were likely a product of
367 the strong effects of IBD in linear distributions, as observed in the conStruct analyses, which
368 suggested a minor contribution of a second layer (K) to explain the genetic structure. These
369 results are in agreement with simulation-based, and empirical studies reporting the high effect of
370 IBD in the linear distribution of river systems, including the formation of differentiated lineages
371 (Selkoe et al., 2015; Thomaz et al., 2016; Farias et al., 2019). However, studies have suggested
372 that the riverway distance plays a minor role in structuring genetic diversity of floodplain
373 populations across the entire Amazonian Basin, suggesting the more pronounced effect of other
374 environmental features (Hrbek et al., 2005; dos Anjos Oliveira et al., 2019; Thom et al., 2020b).
375 Our results reinforce the idea that IBD has to be taken into account when testing for genetic
376 structure in continuous populations on the risk of assuming spurious discrete populations in
377 downstream population genetics analyses.

378

379 *Future directions to explore spatial patterns of genetic diversity across the Amazonian* 380 *floodplains*

381 Our study provided new insights on the spatial dynamics of floodplain forest
382 environments over time, refuting a pattern of common range expansion for co-occurring species.
383 However, our data is limited in the sense that more sampled localities could improve our power
384 in detecting range size changes. Although studies have reported robust results with the
385 rangeExpansion method (Peter & Slatkin, 2013) sampling a single diploid individual for 10
386 localities and 1,500 SNPs, which is a data set similar to ours, the ideal data set lies around 20
387 diploid individuals (1 per locality) and 7,000 SNPs (Peter & Slatkin, 2013, 2015; Potter et al.,
388 2016). An important factor that might hinder our ability to detect if a range expansion occurred is
389 the time since populations reached a stationary condition (Alvarado-Serrano & Hickerson, 2018).
390 All three species we analyzed have extensive distributions across the Solimões River, and might

391 already have reached isolation by distance equilibrium, eroding the signal for directional range
392 expansions. Recent evidence has supported that secondary contact and gene flow are pervasive
393 processes across the floodplains, although controlled by varying degrees of connectivity through
394 time (Thom et al., 2018, 2020b). The methods we applied do not incorporate the effects of gene
395 flow from unsampled populations, which could inflate genetic diversity in eastern Solimões
396 localities, masking the genomic signal of range expansions. The inferred range expansion in *T.*
397 *cryptoleucus* pointed to an interesting scenario, since this taxon is currently in contact with *T.*
398 *nigrocinereus tshudi* from the Madeira River, suggesting diversification in allopatry followed
399 by range expansion and recent secondary contact. These results suggest that the approach we
400 used might be robust to some level of gene flow from distinct populations. Future studies aiming
401 to explore the spatial demographic dynamics of floodplain populations should target widespread
402 geographic distribution of samples (e.g. one individual per locality and many localities), and
403 more genetic markers, obtained with reduced representation techniques such as RADseq
404 (Andrews et al., 2016), or whole-genome sequencing (Ellegren, 2014). Additionally, spatial
405 explicit simulations of demographic histories coupled with supervised machine learning
406 classification algorithms could improve our ability to test range size changes through time with a
407 reduced number of samples ([Battey et al. 2020](#); [Reid et al. 2019](#)).

408 Ecological niche modeling has been used to track the current and paleo distribution of
409 various Amazonian organisms (Prates et al., 2016; dos Anjos Oliveira et al., 2019; Silva et al.,
410 2019). However, the intimate relationship of floodplain species with microhabitats linked to the
411 sedimentation dynamics of the rivers, probably cannot be captured by interpolated bioclimatic
412 variables (e.g. Worldclim). Meaningful variables that could describe the sedimentation process
413 and detailed distribution of floodplain habitats with a good resolution for the entire Basin are
414 becoming available (Pekel et al., 2016; Fassoni-Andrade & Paiva, 2019) and might provide new
415 insights on how the current sediment budget of rivers and vegetation dynamics shape patterns of
416 genetic diversity. Amazonia is currently at the center of the debate about the impact of
417 hydroelectric dams on biodiversity and the sedimentary budget of rivers (Latrubesse et al., 2017).
418 Understanding how the genetic diversity (dos Anjos Oliveira et al., 2019) and species richness
419 (Laranjeiras et al., 2021) might be associated with river dynamism and productivity is paramount
420 to predict future impacts caused by anthropic activities.

421

422 **Acknowledgments**

423 We thank the curatorial staff of the Museu Paraense Emílio Goeldi (MPEG), Instituto Nacional
424 de Pesquisas da Amazônia (INPA), Laboratório de Genética e Evolução Molecular de Aves
425 (LGEMA) for loaning tissue samples under their care. We thank members of the Smith lab,
426 Hickerson lab, and Carnaval lab: Brian Smith, Kaiya L. Provost, Lucas R. Moreira, Vivien Chua,
427 Jon Merwin, Ana C. Carnaval, Connor French, Andrea Paz, Kathryn Mercier, Rilquer
428 Mascarenhas, Lidia Martins, Michael Hickerson, Isaac Overcast, Alexander Xue. We thank L. E.
429 Araujo-Silva for the partnership during field expeditions. We thank Oscar Johnson and Rob
430 Brumfield for sharing UCE sequences. We thank the Research Center on Biodiversity and
431 Computing (BioComp) of the Universidade de São Paulo (USP), supported by the USP Provost's
432 Office for Research. This study was co-funded by FAPESP (BIOTA, 2012/50260-6 and
433 2013/50297-0), NSF (DOB 1343578), NASA, CNPq (310593/2009-3, 574008/2008-0,
434 563236/2010-8, and 471342/2011-4), PEER/USAID (AID-OAA-A-11-00012), and FAPESPA
435 (ICAAF 023/2011). G.T. was granted by CAPES and then FAPESP scholarships (2014/00113-2,
436 2015/12551-7, 2018/17869-3, and 2017/25720-7). A.A., C.C.R., and C.M. are supported by
437 CNPq research productivity fellowships (310880/2012-2, 308927/2016-8, and 3062204/2019-3).

438

439 **Biosketch**

440 **Gregory Thom** is broadly interested in comparative genomics, testing the major factors
441 impacting population differentiation over historical timescales. His overriding goal is to unearth
442 the mechanisms driving Neotropical bird evolution by using genomics and advanced
443 computational biology.

444

445 **Author contributions**

446 G.T. conceived the study and performed bioinformatics and phylogeographic analyses. G.T.,
447 C.C.R., and C.M. wrote the paper with input from A.A. and ES. GT and ES performed the river
448 dynamicity analyses. All authors reviewed and approved the final version of the manuscript
449 before submission.

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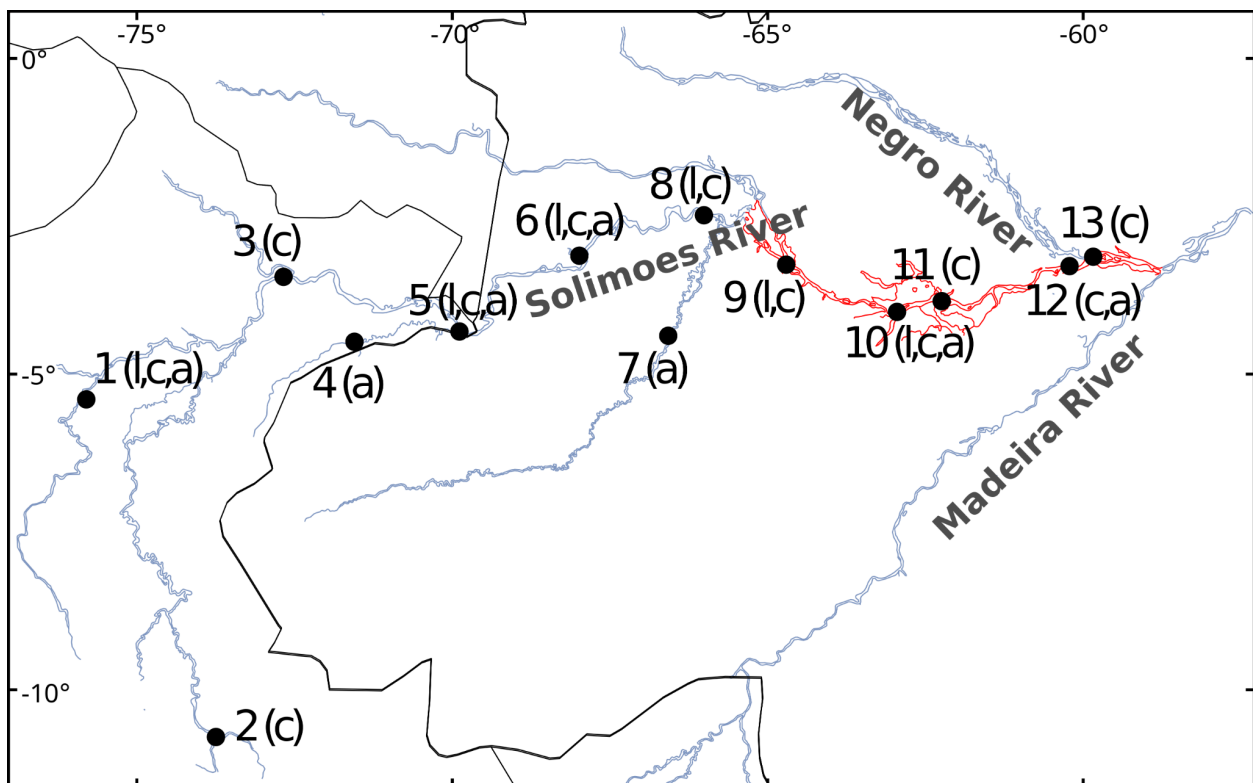
715 Table 1: Summary of results obtained with rangeExpansion for three species of birds occurring
716 across the Solimoes River.

Parameters	<i>M. lugubris</i>	<i>T. cryptoleucus</i>	<i>M. assimilis</i>
longitude	-62.95	-73.22144	-75.64357
latitude	-4.021451	-9.999475	-3148
r1	0.999	0.999	0.999

r10	0.996	0.991	0.997
r100	0.964	0.918	0.975
d1	27.19	11.45	40.80
r ²	0.45	0.89	0.31
p-value	36.17	2.10e-23	11.26

717 r1, r10, and r100 - decrease in diversity at 1, 10, and 100 kilometers of distance, respectively; d1
 718 – effective founder distance, which represents the deme size in km for the effective population
 719 size to be reduced by 1% in the expansion front; r² and p-value - correlation coefficient and
 720 correlation p-value for the most likely origin, respectively.

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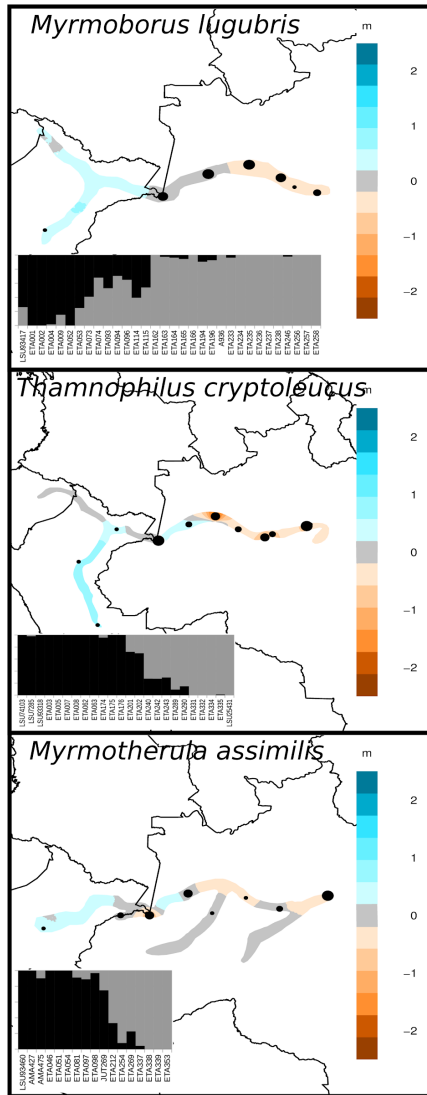


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724 Figure 1: Geographic distribution of sampled individuals of *Myrmoborus lugubris* (l),
 725 *Thamnophilus cryptoleucus* (c), and *Myrmotherula assimilis* (a) across the Solimões River Basin.
 726 Numbers in the map represent localities as in Table S1. In red is the putative zone of unstable

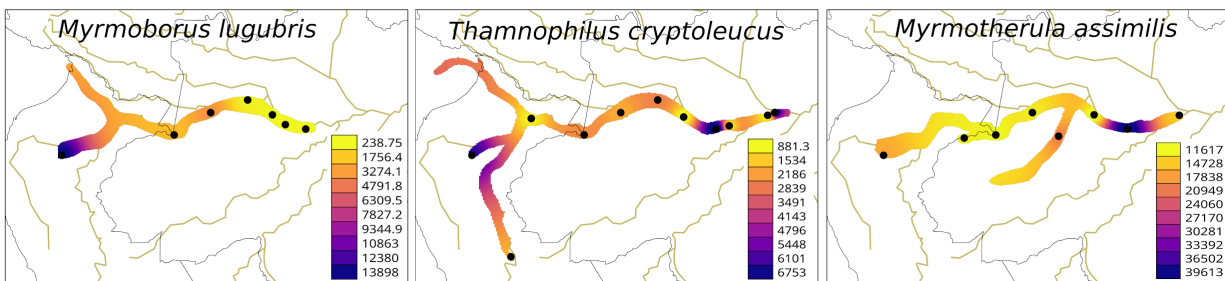
727 habitats following the range expansion hypothesis.



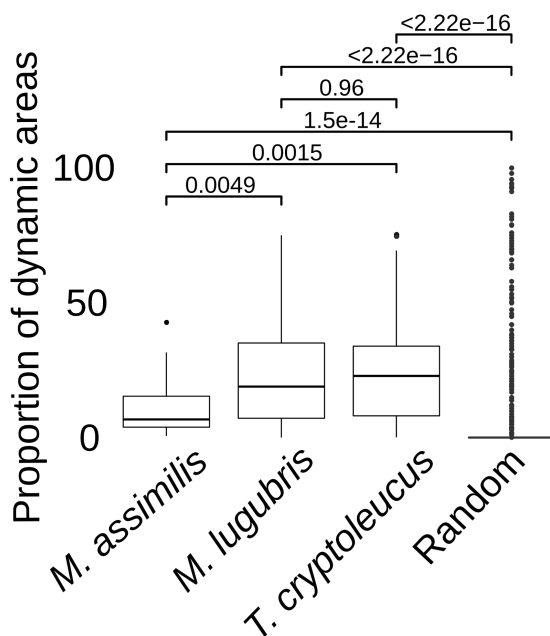
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729 Figure 2: Estimated effective migration surface (EEMS) and population structure and individual
730 coefficient of ancestry (bars) inferred with sNMF for the three species studied. Colors in the map
731 represent a log10 scale from the average effective migration (gray). Codes below the bars in the
732 admixture plot represent identification numbers of individuals as in Table S1. Note that for *M.*
733 *assimilis*, K=2 was the model with the second-highest cross-entropy value.

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 736 Figure 3: Interpolated Harpending's raggedness index estimated from the geographic spectrum
 737 of shared alleles (GSSA) histograms. Lower values represent the most likely source of a serial
 738 range expansion.



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 741 Figure 4: Average river dynamism on 1000 meters radius around randomly drawn points and
 742 occurrence records for *M. assimilis*, *M. lugubris*, and *T. cryptoleucus*. River dynamism was
 743 calculated based on the "Water Occurrence Change" layer available at [https://global-surface-](https://global-surface-water.appspot.com/)
 744 [water.appspot.com/](https://global-surface-water.appspot.com/). Higher values represent higher river dynamicity. The pairwise comparisons
 745 show p-values for Wilcoxon tests between species and random points.

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